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H2FS	0
I2R.USPT,PGPB.	38
I2RS	0
3D1.USPT,PGPB.	165
3D1S	0
ANTIBOD$	0
ANTIBOD.USPT,PGPB.	229
ANTIBODAY.USPT,PGPB.	1
ANTIBODEES.USPT,PGPB.	1
. . . . .	
ANTIBOD$(ANTIBODY/ADA).USPT,PGPB.	pickup term
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Refine Search: `(H2F or I2R or 3D1) same (antibod$ or hybridoma$ or immunoglobulin$)` Clear

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DB Name	Query	Hit Count	Set Name
USPT,PGPB	(H2F or I2R or 3D1) same (antibod$ or hybridoma$ or immunoglobulin$)	8	L1

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Generate Collection

L1: Entry 1 of 8 File: USPT Mar 6, 2001

DOCUMENT-IDENTIFIER: US 6197524 B1
TITLE: Methods for detecting, identifying, isolating, and selectively labelling and targeting TH1 lymphocyte by means of the LAG-3 protein

DEPR:
Detection of soluble LAG-3 in the supernatants of T-cell clones was performed by an appropriate ELISA based on the use of a recombinant soluble LAG-3 derived molecule (sLAG-3D1-D2). sLAG-3D1-D2 was obtained from PCR amplified DNA fragment encoding for the two first immunoglobulin-like domains of LAG-3 and subcloned into the pCLH3AXSV2DHFR expression vector (Cole et al., 1993). The construct was used to transfect a DHFR deficient CHO cell line (DUKX-B11) (Urlaub and Chasin, 1980). The sLAG-3D1-D2-producing CHO cells were cultured in Wheaton bioreactor and sLAG-3D1-D2 molecule purified by capture step on fast SP-Sepharose column followed by immunopurification on a 17B4 mAb-Poros EP column. The resulting protein was found to be >90% pure by RP-HPLC and SDS-PAGE. For the assay, plate wells were coated with anti-LAG-3 (11E3; 10.mu.g/ml on 0.2 M carbonate buffer, pH 9.6) mAb and then incubated for 12 hr with test samples or different dilutions (from 0.12 to 25 ng/ml) of sLAG-3D1-D2. After washings, biotinylated anti-LAG-3 (17B4) mAb (0.5 .mu.g/ml) was added for additional 2 hr, the plates stained with substrate solution, and the reaction read at 492 nm.

L1: Entry 3 of 8 File: USPT Aug 29, 2000

DOCUMENT-IDENTIFIER: US 6110463 A
TITLE: Anti-Cryptosporidium parvum preparations

DEPR:
Hybridomas secreting mAbs listed in the first column of Table 4 were separately propagated in BALB/c mice and ascites fluid samples containing the mAbs collected using standard laboratory methods. These mAb preparations were administered to neonatal BALB/c mice as pools of ascites fluid to test the in vivo neutralizing capacity of the preparations according to the method described under Example 5. The volume of ascites fluid administered to neonatal mice in this procedure was held constant at 150 .mu.l. Equal volumes of different ascites fluid samples were mixed to create preparations of mAb combinations. A pool or collection of mAbs HL113 (IgM), HL245 (IgG.sub.3) and HL296 (IgG.sub.1), all having irrelevant binding specificities, served as a negative control in the procedure. A second pool included mAbs 18.44, C6B6, C4A1, 2B4, 3D1, 3E2 and M23A1, all having binding specificities for epitopes disposed on C. parvum. In this procedure, 8-10 mice were used for each test group.

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Term	Documents
H2F.USPT,PGPB.	8
H2FS	0
I2R.USPT,PGPB.	38
I2RS	0
3D1.USPT,PGPB.	165
3D1S	0
CHIMERIC.USPT,PGPB.	10178
CHIMERICS.USPT,PGPB.	60
CHIMAERIC.USPT,PGPB.	592
CHIMAERICS.USPT,PGPB.	2
((H2F OR I2R OR 3D1) SAME (CHIMERIC OR CHIMAERIC OR HUMANIZ$ OR HUMANIS$)).USPT,PGPB.	0

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Search History

Today's Date: 6/23/2001

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USPT,PGPB	(H2F or I2R or 3D1) same (chimeric or chimaeric or humaniz$ or humanis$)	0	L2
USPT,PGPB	(H2F or I2R or 3D1) same (antibod$ or hybridoma$ or immunoglobulin$)	8	L1